

Technology Review

Atomic Absorption Spectroscopy in Ion Channel Screening

Larisa Stankovich, Sasko Despotovski, and Dong Liang

Abstract: This article examines the utility of atomic absorption spectroscopy, in conjunction with cold flux assays, to ion channel screening. The multiplicity of ion channels that can be interrogated using cold flux assays and atomic absorption spectroscopy is summarized. The importance of atomic absorption spectroscopy as a screening tool is further elaborated upon by providing examples of the relevance of ion channels to various physiological processes and targeted diseases.

Towards Ion Channel Drug Discovery and Development

RECENT YEARS HAVE SEEN a significant shift of resources in the pharmaceutical industry towards the area of ion channel drug discovery and development. This shift was compelled by the issue of drug-induced QT prolongation and the emergence of the S7B, E14 guidance documents for drug safety assessment. The importance of the hERG ion channel to drug development created significant interest in other ion channel targets. As a result, over 6 billion dollars in yearly sales and 15% of the top-selling drugs are targeted for ion channels.¹⁻³

Physiological Significance of Ion Channels

Ion channels are integral membrane proteins that permit the flow of ions, such as calcium, potassium, sodium, and chloride, into and out of cells. Ion channels are present in all human cells and are involved in functions such as nerve transmission, cellular homeostasis, hormonal secretion, and the heartbeat. The wide range of physiological processes in which ion channels are involved is suggested by Table 1.

Channelopathies

With ion channels playing such an important role in many physiological functions, it is not surprising that ion channel dysfunction has been implicated in a number of diseases and disorders. Diseases that are caused by defective ion channel proteins are termed “channelopathies.” Indeed, ion channels represent one of today’s more promising and exciting classes of therapeutic targets on account of the broad range of conditions that are poised to benefit from agents that modulate ion channel activity. Table 2 lists some of the diseases and disorders in which ion channel dysfunction has been implicated.

Flux Assays in Ion Channel Screening

A number of technologies are available to screen against ion channel targets. Xu *et al.*⁴⁶ segmented the ion channel assay market and identified the advantages and disadvantages of various technologies. Available technologies include manual patch-clamp, automated patch-clamp instruments, fluorescence-based platforms, membrane binding assays, radioactive ionic flux assays, and non-radioactive flux assays. However, this review

T1

T2

TABLE 1. PHYSIOLOGICAL PROCESSES INVOLVING ION CHANNELS

<i>Physiological process</i>	<i>Ion channel or gene encoding ion channel</i>
Apoptosis	MAC ⁴
Bicarbonate secretion	Purinergic P2X ⁵
Bone resorption	SLC26 ⁶
	CIC-7 ⁷
	RyR ⁸
Cardiac repolarization	KCNQ1, hERG, SCN5A, KCNE1, KCNE2 ⁹
Chloride absorption	CFTR ¹⁰
Inflammation	ASIC's ¹¹
Insulin secretion	K _{ATP} (Kir6.2 and SUR1 subunits) ¹²
Metastasis	CLCA ¹³
Neuronal excitability	KCNQ2/KCNQ3 ¹⁴
	KCNQ5 ¹⁵
	T-type Ca channels ¹⁶
Neuronal migration	Kv4.3 ¹⁷
Nociception	L-type Ca channels ¹⁸
Regulation of basal metabolic rate	Kv1.3 ¹⁹
Smooth muscle relaxation	
Cavernosal	BK _{Ca} ²⁰
Urinary	SK _{Ca} , BK _{Ca} , and RyR ²¹
Vascular	KCNMB1 ²²
Sperm motility	PKD2 ²³
T cell activation	Kv1.1, Kv1.3 ²⁴

TABLE 2. DISEASES OR DISORDERS INVOLVING ION CHANNELS

<i>Disease or disorder</i>	<i>Ion channel or gene encoding ion channel</i>
Andersen-Tawil's syndrome	KCNJ2 ²⁵
Bartter's syndrome	SLC12A2, KCNJ1, ROMK, CIC-Ka, CICKb ²⁶
Benign familial neonatal convulsions	KCNQ2, KCN3 ¹⁴
	SCN2A ²⁷
Brody's disease	SERCA1 ²⁸
Brugada's syndrome	SCN5A ²⁹
Periodic paralysis	SCN4A ^{30,31}
Cancer	CLCA ¹³
Cystic fibrosis	CFTR ¹⁰
	SLC26 ⁶
Epilepsy	T-type Ca channels ¹⁶
Erectile dysfunction	BK _{Ca} ²⁰
Familial hemiplegic migraine	CACNA1A ³²
Heart failure	Cardiac P2X4 ³³
Hypertension	KCNMB1 ²²
Isolated cardiac conduction disease	SCN5A ³⁴
Incontinence	SK _{Ca} , BK _{Ca} , and RyR ²¹
Inflammation	ASIC's ¹¹
Lenegre's disease	SCN5A ³⁵
Long-QT syndrome	KCNQ1, hERG, SCN5A, KCNE1, KCNE2 ⁹
	HCN4 ³⁶
Male sterility	PKD2 ²³
Multiple sclerosis	Kv1.3 ³⁷
	Nav1.2, Nav1.6, Na ⁺ /Ca ²⁺ exchanger ³⁸
Myotonia congenita	CLCN1 ^{39,40}
Myotonic dystrophy	SERCA1 ²⁸
Neuropathic pain	Nav1.8 ⁴¹
	Nav1.9 ⁴²
Obesity	Kv1.3 ¹⁹
Osteopetrosis	CIC-7 ⁷
Polycystic kidney disease	PKD2 ⁴³
Schizophrenia	SK3 ⁴⁴
Type 1 diabetes	K _{ATP} (Kir6.2 subunit) ⁴⁵
Type 2 diabetes	K _{ATP} (Kir6.2 and SUR1 subunits) ¹²

TABLE 3. CHANNELS THAT HAVE BEEN EXAMINED USING THE COLD FLUX ASSAY APPROACH

Family	Ion channel or gene encoding ion channel
K	hERG ^{49,50} KCNQ2 ⁵¹ BK _{Ca} ^{48,52} SK _{Ca} ⁴⁸ Kv1.1, Kv1.4 ⁴⁸ Kv1.3 ⁵³
Na	SCN5A ⁵⁴
Cl	CFTR ⁵⁵
Ca	Cardiac L-type ⁵⁶
Non-selective cation	Nicotinic acetylcholine ⁴⁸ Purinergic P2X ⁴⁸
Transporter	Na ⁺ /K ⁺ pump ⁵⁷ K ⁺ -Cl ⁻ co-transporter ⁵⁸

will focus on the non-radioactive (cold) flux assay technology from Aurora Biomed Inc. (Vancouver, BC, Canada).

Non-radioactive flux assay technology utilizes highly sensitive flame atomic absorption spectrometers capable of dealing with the low sample volumes that are typically used in pharmaceutical research. Cold flux assays use tracer ions: molecules that are similar in size and charge to the ion of interest. To study potassium and sodium channels, the rubidium and lithium ions, respectively, are used. For chloride channels, the chloride ion itself is used with an indirect detection procedure involving precipitation with silver nitrate and measurement of the concentration of free silver.

The use of flux assays in high-throughput screening of ion channels for pharmaceutical research has been well summarized by Gill *et al.*⁴⁷ The cold flux assay as applied to the study of ion channel function is a relatively novel approach and was first proposed by Terstappen.⁴⁸ Since then, the use of cold flux assays in ion channel

screening has steadily increased, and publications describing the various channels examined are listed in Table 3.

Hot flux (radioactive) assays, on the other hand, have been used for many years to assess ion channel function. Hot flux assays employ radiotracers to monitor ion flux through channels. Some of the ion channels that have been examined using radiotracers are listed in Table 4. Overall, Tables 3 and 4 represent some of the most recent and significant literature on their respective topics.

Any of the channels studied using the hot flux assay approach should, in theory, be amenable to the cold flux assay approach. The cold flux assays that have been developed for the Ion Channel Readers by the Aurora Biomed team include the Rb flux assay against the hERG⁵⁰ and Kv1.3⁵⁶ channels as well as against the Na⁺/K⁺ pump⁵⁷ and K⁺-Cl⁻ co-transporter⁵⁸; the Li flux assay against the SCN5A channel⁵³; and the Cl flux assay for the CFTR channel.⁵⁴ The multiplicity of ion channels that

T3

T4

TABLE 4. CHANNELS THAT HAVE BEEN EXAMINED USING THE HOT FLUX ASSAY APPROACH

Family	Ion channel or gene encoding ion channel	Radiotracer
K	hERG ⁵⁹ BK _{Ca} ⁶⁰ IK _{Ca} ^{61,62} SK _{Ca} ⁶³ Kv ⁶³ KCh ⁶³	⁸⁶ Rb ⁺ ⁸⁶ Rb ⁺ ⁸⁶ Rb ⁺ ⁸⁶ Rb ⁺ ⁸⁶ Rb ⁺ ⁸⁶ Rb ⁺
Na	Neuronal Na _v ⁶⁴	²² Na ⁺
Cl	CFTR ⁶⁵	¹²⁵ I ⁻
Ca	Neuronal Ca _v ⁶⁶	⁴⁵ Ca ²⁺
Non-selective cation	Nicotinic acetylcholine ^{67,68} Purinergic P2X ⁶⁹	⁸⁶ Rb ⁺ ⁸⁶ Rb ⁺
Transporter	Na ⁺ /K ⁺ pump ⁷⁰ K ⁺ -Cl ⁻ co-transporter ^{71,72} Na ⁺ -K ⁺ -Cl ⁻ co-transporter ^{73,74}	⁸⁶ Rb ⁺ ⁸⁶ Rb ⁺ ⁸⁶ Rb ⁺ , ¹²⁵ I ⁻

can be examined using the cold flux assay approach is indicative of the versatility of this technique for ion channel screening.

Acknowledgments

The authors are grateful to David Wicks for his help in the revision process.

References

- Augustin HG: Antiangiogenic tumour therapy: will it work? *Trends Pharmacol Sci* 1998;19:216–222.
- Baxter DF, Kirk M, Garcia AF, Raimondi A, Holmqvist MH, Flint KK, Bojanic D, Distefano PS, Curtis R, Xie Y: A novel membrane potential-sensitive fluorescent dye improves cell-based assays for ion channels. *J Biomol Screen* 2001;7:79–85.
- Berman FW, Murray TF: Brevetoxin-induced autocrine excitotoxicity is associated with manifold routes of Ca^{2+} influx. *J Neurochem* 2000;74:1443–1451.
- Guo L, Pietkiewicz D, Pavlov EV, Grigoriev SM, Kasianowicz JJ, Dejean LM, Korsmeyer SJ, Antonsson B, Kinnally KW: Effects of cytochrome c on the mitochondrial apoptosis-induced channel MAC. *Am J Physiol Cell Physiol* 2003;286:C1109–C1117.
- Sellick GS, Rudd M, Eve P, Allinson R, Matutes E, Catovsky D, Houlston RS: The P2X7 receptor gene A1513C polymorphism does not contribute to risk of familial or sporadic chronic lymphocytic leukemia. *Cancer Epidemiol Biomarkers Prev* 2004;13:1065–1067.
- Ko SB, Zeng W, Dorwart MR, Luo X, Kim KH, Millen L, Goto H, Naruse S, Soyombo A, Thomas PJ, Muallem S: Gating of CFTR by the STAS domain of SLC26 transporters. *Nat Cell Biol* 2004;6:343–350.
- Henriksen K, Gram J, Schaller S, Dahl BH, Dziegiel MH, Bollerslev J, Karsdal MA: Characterization of osteoclasts from patients harboring a G215R mutation in *CLC-7* causing autosomal dominant osteopetrosis type II. *Am J Pathol* 2004;164:1537–1545.
- Zaidi M, Moonga BS, Huang CL: Calcium sensing and cell signaling processes in the local regulation of osteoclastic bone resorption. *Biol Rev Camb Philos Soc* 2004;79:79–100.
- Fodstad H, Swan H, Laitinen P, Piippo K, Paavonen K, Viitasalo M, Toivonen L, Kontula K: Four potassium channel mutations account for 73% of the genetic spectrum underlying long-QT syndrome (LQTS) and provide evidence for a strong founder effect in Finland. *Ann Med* 2004;36:53–63.
- Cornet-Boyaka E, Jablonsky M, Naren AP, Jackson PL, Muccio DD, Kirk KL: Rescuing cystic fibrosis transmembrane conductance regulator (CFTR)-processing mutants by transcomplementation. *Proc Natl Acad Sci USA* 2004;101:8221–8226.
- Mamet J, Baron A, Lazdunski M, Nicolas V: Proinflammatory mediators, stimulators of sensory neuron excitability via the expression of acid-sensing ion channels. *J Neurosci* 2002;22:10662–10670.
- Florez JC, Burt N, de Bakker PI, Almgren P, Tuomi T, Holmkvist J, Gaudet D, Hudson TJ, Schaffner SF, Daly MJ, Hirschhorn JN, Groop L, Altshuler N: Haplotype structure and genotype-phenotype correlations of the sulfonylurea receptor and the islet ATP-sensitive potassium channel gene region. *Diabetes* 2004;53:1360–1368.
- Abdel-Ghany M, Cheng HC, Elble RC, Lin H, DiBiasio J, Pauli BU: The interacting binding domains of the beta(4) integrin and calcium-activated chloride channels (CLCAs) in metastasis. *J Biol Chem* 2003;278:49406–49416.
- Singh NA, Westenskow P, Charlier C, Pappas C, Leslie J, Dillon J, Anderson VE, Sanguinetti MC, Leppert MF, BFNC Physician Consortium: KCNQ2 and KCNQ3 potassium channel genes in benign familial neonatal convulsions: expansion of the functional and mutation spectrum. *Brain* 2003;126:2726–2737.
- Schroeder BC, Hechenberger M, Weinreich F, Kubisch C, Jentsch TJ: KCNQ5, a novel potassium channel broadly expressed in brain, mediates M-type currents. *J Biol Chem* 2000;275:24089–24095.
- Lacinova L: Pharmacology of recombinant low-voltage activated calcium channels. *Curr Drug Targets* 2004;3:105–111.
- Hsu YH, Huang HY, Tsaor ML: Contrasting expression of Kv4.3, an A-type K^+ channel, in migrating Purkinje cells and other post-migratory cerebellar neurons. *Eur J Neurosci* 2003;18:601–612.
- Todorovic SM, Pathirathna S, Meyenburg A, Jevtovic-Todorovic V: Mechanical and thermal anti-nociception in rats after systemic administration of verapamil. *Neurosci Lett* 2004;360:57–60.
- Xu J, Koni PA, Wang P, Li G, Kaczmarek L, Wu Y, Li Y, Flavell RA, Desir GV: The voltage-gated potassium channel Kv1.3 regulates energy homeostasis and body weight. *Hum Mol Genet* 2003;12:551–559.
- Archer SL: Potassium channels and erectile dysfunction. *Vascul Pharmacol* 2002;38:61–71.
- Heppner TJ, Herrera GM, Bonev AD, Hill-Eubanks D, Nelson MT: Ca^{2+} sparks and K_{Ca} channels: novel mechanisms to relax urinary bladder smooth muscle. *Adv Exp Med Biol* 2003;539:347–357.
- Fernandez-Fernandez JM, Tomas M, Vazquez E, Orío P, Latorre R, Senti M, Marrugat J, Valverde MA: Gain-of-function mutation in the KCNB1 potassium channel subunit is associated with low prevalence of diastolic hypertension. *J Clin Invest* 2004;113:1032–1039.
- Gao Z, Ruden DM, Lu X: PKD2 cation channel is required for directional sperm movement and male fertility. *Curr Biol* 2003;13:2175–2178.
- Beeton C, Barbaria J, Giraud P, Devaux J, Benoliel AM, Gola M, Sabatier JM, Bernard D, Crest M, Beraud E: Selective blocking of voltage-gated K^+ channels improves experimental autoimmune encephalomyelitis and inhibits T cell activation. *J Immunol* 2001;166:936–944.
- Donaldson MR, Yoon G, Fu YH, Ptacek LJ: Andersen-Tawil syndrome: a model of clinical variability, pleiotropy, and genetic heterogeneity. *Ann Med* 2004;36:92–97.
- Hebert SC: Bartter syndrome. *Curr Opin Nephrol Hypertens* 2003;12:527–532.
- Berkovic S, Heron SE, Giordano L, Marini C, Guerrini R, Kaplan RE, Gambardella A, Steinlein OK, Grinton BE, Dean JT, Bordo L, Hodgson BL, Yamamoto T, Mulley JC, Zara F, Scheffer IE: Benign familial neonatal-infantile seizures: characterization of a new sodium channelopathy. *Ann Neurol* 2004;55:550–557.
- Benders AA, Wevers RA, Veerkamp JH: Ion transport in human skeletal muscle cells: disturbances in myotonic dystrophy and Brody's disease. *Acta Physiol Scand* 1996;156:355–367.
- Baroudi G, Napolitano C, Priori SG, Del Bufalo A, Chahine

- M: Loss of function associated with novel mutations of the SCN5A gene in patients with Brugada syndrome. *Can J Cardiol* 2004;20:425–430.
30. Brancati F, Valente EM, Davies NP, Sarkozy A, Sweeney MG, LoMonaco M, Pizzuti A, Hanna MG, Dallapiccola B: Severe infantile hyperkalaemic periodic paralysis and paramyotonia congenita: broadening the clinical spectrum associated with the T704M mutation in SCN4A. *J Neurol Neurosurg Psychiatry* 2003;74:1339–1341.
 31. Sugiura Y, Makita H, Li L, Noble PJ, Kimura J, Kumagai Y, Soeda T, Yamamoto T: Cold induces shifts of voltage dependence in mutant SCN4A, causing hypokalemic periodic paralysis. *Neurology* 2003;14:914–918.
 32. Akerman S, Williamson DJ, Goadsby PJ: Voltage-dependent calcium channels are involved in neurogenic dural vasodilation via a presynaptic transmitter release mechanism. *Br J Pharmacol* 2003;3:558–566.
 33. Yang A, Sonin D, Jones L, Barry WH, Liang BT: A beneficial role of cardiac P2X₄ receptors in heart failure: rescue of the calyculin A overexpression model of cardiomyopathy. *Am J Physiol Heart Circ Physiol* 2004;287:H1096–H1103. E pub 2004 May 6.
 34. Tan HL, Bink-Boelkens MT, Bezzina CR, Viswanthan PC, Beaufort-Krol GC, van Tintelen PJ, van den Berg MP, Wilde AA, Balser JR: A sodium channel mutation causes isolated cardiac conduction disease. *Nature* 2001;409:1043–1047.
 35. Probst V, Kyndt F, Potet F, Trochu JN, Mialet G, Demolombe S, Schot JJ, Baro I, Escande D, Le Marec H: Haploinsufficiency in combination with aging causes SCN5A-linked hereditary Lenegre disease. *J Am Coll Cardiol* 2003;4:643–652.
 36. Ueda K, Nakamura K, Hayashi T, Inagaki N, Takahashi M, Arimura T, Morita H, Higashiesato Y, Hirano Y, Yasunami M, Takishita S, Yamashina A, Ohe T, Sunamori M, Hiraoka M, Kimura A: Functional characterization of a trafficking-defective HCN4 mutation, D553N, associated with cardiac arrhythmia. *J Biol Chem* 2004;279:27194–27198. E pub 2004 Apr 30.
 37. Wulff H, Calabresi PA, Allie R, Yun S, Pennington M, Beeton C, Chandy KG: The voltage-gated Kv1.3 K⁺ channel in effector memory T cells as new target for MS. *J Clin Invest* 2003;112:298.
 38. Craner MJ, Newcombe J, Black JA, Hartle C, Cuzner ML, Waxman SG: Molecular changes in neurons in multiple sclerosis: altered axonal expression of Nav1.2 and Nav1.6 sodium channels and Na⁺/Ca²⁺ exchanger. *Proc Natl Acad Sci USA* 2004;101:8168–8173.
 39. Chen L, Schaerer M, Lu ZH, Lang D, Joncourt F, Weis J, Fritsch J, Kappeler L, Gallati S, Sigel E, Burgunder JM: Exon 17 skipping in CLCN1 leads to recessive myotonia congenita. *Muscle Nerve* 2004;29:670–676.
 40. Grunnet M, Jespersen T, Colding-Jorgensen E, Schwartz M, Klaerke DA, Vissing J, Olesen SP, Duno M: Characterization of two new dominant CIC-1 channel mutations associated with myotonia. *Muscle Nerve* 2003;28:722–732.
 41. Roza C, Laird JM, Souslova V, Wood JN, Cervero F: The tetrodotoxin-resistant Na⁺ channel Nav1.8 is essential for the expression of spontaneous activity in damaged sensory axons of mice. *J Physiol* 2003;550:921–926.
 42. Coste B, Osorio N, Padilla F, Crest M, Delmas P: Gating and modulation of presumptive Nav1.9 channels in enteric and spinal sensory neurons. *Mol Cell Neurosci* 2004;26:123–132.
 43. Stekrova J, Reiterova J, Merta M, Damborsky J, Zidovska J, Kebrdlova V, Kohoutova M: PKD2 mutations in a Czech population with autosomal dominant polycystic kidney disease. *Nephrol Dial Transplant* 2004;19:1116–1122.
 44. Tomita H, Shakkottai VG, Gutman GA, Sun G, Bunney WE, Cahalan MD, Chandy KG, Gargus JJ: Novel truncated isoform of SK3 potassium channel is a potent dominant-negative regulator of SK currents: implications in schizophrenia. *Mol Psychiatry* 2003;8:524–535, 460. Erratum in: *Mol Psychiatry* 2003;8:766.
 45. Gloyd AL, Pearson ER, Antcliff JF, Proks P, Bruining GJ, Slingerland AS, Howard N, Srinivasan S, Silva JM, Molnes J, Edghill EL, Frayling TM, Temple IK, Mackay D, Shield JP, Sumnik Z, van Rhijn A, Wales JK, Clark P, Gorman S, Aisenberg J, Ellard S, Njolstad PR, Ashcroft FM, Hattersley AT: Activating mutations in the gene encoding the ATP-sensitive potassium-channel subunit Kir6.2 and permanent neonatal diabetes. *N Engl J Med* 2004;350:1838–1849.
 46. Xu J, Wang X, Ensign B, Li M, Guida A, Xu J: Ion channel assay technologies: quo vadis? *Drug Discov Today* 2001;6:1278–1287.
 47. Gill S, Gill R, Lee SS, Hesketh JC, Fedida D, Rezazadeh S, Stankovich L, Liang D: Flux assays in high throughput screening of ion channels in drug discovery. *Assay Drug Dev Technol* 2003;1:709–717.
 48. Terstappen GC: Functional analysis of native and recombinant ion channels using a high capacity non-radioactive rubidium efflux assay. *Anal Biochem* 1999;272:149–155.
 49. Tang W, Kang J, Wu X, Rampe D, Wang L, Shen H, Li Z, Dunnington D, Garyantes T: Development and evaluation of high throughput functional assay methods for hERG potassium channel. *J Biomol Screen* 2001;6:325–331.
 50. Aurora Biomed Laboratory: *Validation Report: Using the HEK-hERG cell Line with Aurora Biomed's Ion Channel Reader*. Aurora Biomed Inc., Vancouver, BC, Canada, 2002.
 51. Scott CW, Wilkins DE, Trivedi S, Crankshaw DJ: A medium-throughput functional assay of KCNQ2 potassium channels using rubidium efflux and atomic absorption spectrometry. *Anal Biochem* 2003;319:251–257.
 52. Parihar AS, Groebe DR, Scott VE, Feng J, Zhang XF, Warrior U, Gopalakrishnan M, Shieh CC: Functional analysis of large conductance Ca²⁺-activated K⁺ channels: ion flux studies by atomic absorption spectrometry. *Assay Drug Dev Technol* 2003;5:647–654.
 53. Aurora Biomed Laboratory: *Preliminary Report on Development of HTS Assay for Kv1.3 Channel*. Aurora Biomed Inc., Vancouver, BC, Canada, 2004.
 54. Aurora Biomed Laboratory: *Preliminary Report on the Development of a HTS Assay for SCN5A Sodium Channel*. Aurora Biomed Inc., Vancouver, BC, Canada, 2003.
 55. Aurora Biomed Laboratory: *Preliminary Report on the Development of a HTS Assay for CFTR Channel and ICR*. Aurora Biomed Inc., Vancouver, BC, Canada, 2003.
 56. Walsh KB, Cheng Q: Intracellular Ca(2+) regulates responsiveness of cardiac L-type Ca(2+) current to protein kinase A: role of calmodulin. *Am J Physiol Heart Circ Physiol* 2004;286:H186–H194.
 57. Aurora Biomed Laboratory: *Preliminary Report on the Development of a HTS Assay Na⁺-K⁺-ATPase using ICR*. Aurora Biomed Inc., Vancouver, BC, Canada, 2003.
 58. Aurora Biomed Laboratory: *Preliminary Report on Development of HTS Assay for K⁺-Cl⁻ Co-Transporter*. Aurora Biomed Inc., Vancouver, BC, Canada, 2003.
 59. Cheng CS, Alderman D, Kwash J, Dessaint J, Patel R, Lescoe MK, Kinrade MB, Yu W: A high-throughput HERG potassium channel function assay: an old assay with a new look. *Drug Dev Ind Pharm* 2002;28:177–191.

60. Ahluwalia J, Tinker A, Clapp LH, Duchon MR, Abramov AY, Pope S, Nobles M, Segal AW: The large-conductance Ca^{2+} -activated K^{+} channel is essential for innate immunity. *Nature* 2004;427:853–858.
61. Parihar AS, Coghlan MJ, Gopalakrishnan M, Shieh CC: Effects of intermediate-conductance Ca^{2+} -activated K^{+} channel modulators on human prostate cancer cell proliferation. *Eur J Pharmacol* 2003;471:157–164.
62. de-Allie FA, Bolsolver SR, Nowicky AV, Strong PN: Characterization of Ca^{2+} -activated $^{86}\text{Rb}^{+}$ fluxes in rat C6 glioma cells: a system for identifying novel IKCa-channel toxin. *Br J Pharmacol* 1997;117:479–487.
63. de Silva HA, Carver JG, Aronson JK: Pharmacological evidence of calcium-activated and voltage-gated potassium channels in human platelets. *Clin Sci (Lond)* 1997;93:249–255.
64. Nicholson GM, Blanche T, Mansfield K, Tran Y: Differential blockade of neuronal voltage-gated Na^{+} and K^{+} channels by antidepressant drugs. *Eur J Pharmacol* 2002;452:35–48.
65. Becq F, Auzanneau C, Norez C, Derand R, Bulteau-Pignoux L: Radiotracer flux method to study CFTR channel activity: regulation, pharmacology and drug discovery. The European Working Group on CFTR Expression, 2003.
66. de los Rios C, Marco JL, Carreiras MD, Chinchon PM, Garcia AG, Villarroja M: Novel tacrine derivatives that block neuronal calcium channels. *Bioorg Med Chem* 2002;10:2077–2088.
67. Marks MJ, Whiteaker P, Grady SR, Picciotto MR, McIntosh JM, Collins AC: Characterization of [(125)I]epibatidine binding and nicotinic agonist mediated $^{86}\text{Rb}^{+}$ efflux in interpeduncular nucleus and inferior colliculus of beta2 null mutant mice. *J Neurochem* 2002;81:1102–1115.
68. Avila AM, Davila-Garcia MI, Ascarrunz VS, Xiao Y, Keller KJ: Differential regulation of nicotinic acetylcholine receptors in PC12 cells by nicotine and nerve growth factor. *Mol Pharmacol* 2003;64:974–986.
69. Petit P, Hillaire-Buys D, Manteghetti M, Debrus S, Chapal J, Loubatieres-Mariani MM: Evidence for two different types of P2 receptors stimulating insulin secretion from pancreatic B cell. *Br J Pharmacol* 1998;125:1368–1374.
70. Prasanna G, Dibas A, Hulet C, Yorio T: Inhibition of $\text{Na}^{+}/\text{K}^{+}$ ATPase by endothelin-1 in human nonpigmented ciliary epithelial cells. *J Pharmacol Exp Ther* 2001;296:966–971.
71. Kelley SJ, Thomas R, Dunham PB: Candidate inhibitor of the volume-sensitive kinase regulating K-Cl cotransport: the myosin light chain kinase inhibitor ML-7. *J Membr Biol* 2000;178:31–41.
72. Payne JA: Functional characterization of the neuronal-specific K-Cl cotransporter: implications for $[\text{K}^{+}]_0$ regulation. *Am J Physiol* 1997;273:C1516–C1525.
73. Gillen CM, Forbush F: Functional interaction of the K-Cl cotransporter (KCC1) with the Na-K-Cl cotransporter in HEK-293 cells. *Am J Physiol* 1999;276:C328–C336.
74. Ferrandi M, Salardi S, Parenti P, Ferrari P, Bianchi G, Braw R, Karlisch SJ: $\text{Na}^{+}/\text{K}^{+}/\text{Cl}^{-}$ -cotransporter mediated Rb^{+} fluxes in membrane vesicles from kidneys of normotensive and hypertensive rats. *Biochim Biophys Acta* 1990;1021:13–20.

Address reprint requests to:

Larisa Stankovich

Aurora Biomed Inc.

1001 East Pender Street

Vancouver, BC, Canada V6A 1W2

E-mail: larisa@aurorabiomed.com

STANKOVICH

AU1

References renumbered by order of citation

AU2

Specify presented at meeting, or give publication data